

RESEARCH ARTICLE

Open Access



Development of a bedside tool to predict the probability of drug-resistant pathogens among hospitalized adult patients with gram-negative infections

Thomas P. Lodise^{1*}, Nicole Gidaya Bonine², Jiatao Michael Ye³, Henry J. Folse⁴ and Patrick Gillard²

Abstract

Background: We developed a clinical bedside tool to simultaneously estimate the probabilities of third-generation cephalosporin-resistant Enterobacteriaceae (3GC-R), carbapenem-resistant Enterobacteriaceae (CRE), and multidrug-resistant *Pseudomonas aeruginosa* (MDRP) among hospitalized adult patients with Gram-negative infections.

Methods: Data were obtained from a retrospective observational study of the Premier Hospital that included hospitalized adult patients with a complicated urinary tract infection (cUTI), complicated intra-abdominal infection (cIAI), hospital-acquired/ventilator-associated pneumonia (HAP/VAP), or bloodstream infection (BSI) due to Gram-negative bacteria between 2011 and 2015. Risk factors for 3GC-R, CRE, and MDRP were ascertained by multivariate logistic regression, and separate models were developed for patients with community-acquired versus hospital-acquired infections for each resistance phenotype ($N = 6$). Models were converted to a singular user-friendly interface to estimate the probabilities of a patient having an infection due to 3GC-R, CRE, or MDRP when ≥ 1 risk factor was present.

Results: Overall, 124,068 patients contributed to the dataset. Percentages of patients admitted for cUTI, cIAI, HAP/VAP, and BSI were 61.6, 4.6, 16.5, and 26.4%, respectively (some patients contributed > 1 infection type). Resistant infection rates were 1.90% for CRE, 12.09% for 3GC-R, and 3.91% for MDRP. A greater percentage of the resistant infections were community-acquired relative to hospital-acquired (CRE, 1.30% vs 0.62% of 1.90%; 3GC-R, 9.27% vs 3.42% of 12.09%; MDRP, 2.39% vs 1.59% of 3.91%). The most important predictors of having an 3GC-R, CRE or MDRP infection were prior number of antibiotics; infection site; infection during the previous 3 months; and hospital prevalence of 3GC-R, CRE, or MDRP. To enable application of the six predictive multivariate logistic regression models to real-world clinical practice, we developed a user-friendly interface that estimates the risk of 3GC-R, CRE, and MDRP simultaneously in a given patient with a Gram-negative infection based on their risk (Additional file 1).

Conclusions: We developed a clinical prediction tool to estimate the probabilities of 3GC-R, CRE, and MDRP among hospitalized adult patients with confirmed community- and hospital-acquired Gram-negative infections. Our predictive model has been implemented as a user-friendly bedside tool for use by clinicians/healthcare professionals to predict the probability of resistant infections in individual patients, to guide early appropriate therapy.

Keywords: Antimicrobial resistance, Gram-negative Bacteria, Prediction model

* Correspondence: thomas.lodise@acphs.edu

¹Albany College of Pharmacy and Health Sciences, Albany, NY 12208-3492, USA

Full list of author information is available at the end of the article



Background

The prevalence of highly resistant Gram-negative infections has increased dramatically over the past decade among hospitalized patients throughout the United States [1] and the world [2]. Three of the more concerning antibiotic-resistant Gram-negative pathogens that are classified as serious or urgent threats to public health include third-generation cephalosporin-resistant Enterobacteriaceae (3GC-R), carbapenem-resistant Enterobacteriaceae (CRE), and multidrug-resistant *Pseudomonas aeruginosa* (MDRP) [1, 2].

Patients with infections due to 3GC-R, CRE, or MDRP have limited treatment options and are highly vulnerable to receiving inappropriate empiric or delayed appropriate therapy because clinicians frequently fail to recognize these infections prior to culture and susceptibility reporting. The deleterious consequences of delayed appropriate therapy are well described [3–10]. Not only does delayed appropriate therapy increase the risk of death [3, 5–10], but studies have also shown that delayed appropriate therapy prolongs hospitalization [3–10]. Prolonged hospitalization places patients at risk of developing subsequent antibiotic-resistant infections [3, 11], which can ultimately lead to further antibiotic usage [5] and exacerbate institutional antimicrobial resistance patterns [12, 13].

As a mechanism to promote appropriate antibiotic usage within a healthcare institution, the World Health Organization recommends that healthcare institutions create tools and implement policies informed by real-world data to increase the probability of patients receiving early appropriate therapy [14]. To enable the administration of early and appropriate therapy, it is important that patients at risk of being infected with resistant pathogens be identified promptly, especially as definitive culture results are typically not available within the first 24 to 72 h of infection onset [15, 16]. Although there is a general appreciation of patient-level risk factors (eg, comorbid conditions, immunosuppression) and exposures (eg, cumulative number of prior antibiotic exposures, prior hospitalizations) that increase the probability of having an infection due to 3GC-R, CRE, or MDRP, there are no widely available and readily adaptable tools for estimating the likelihood of having one of these resistant Gram-negative pathogens simultaneously in individual patients according to the presence or absence of critical hospital- and patient-level covariates.

To address this unmet need, the goal of this study was to develop a user-friendly clinical tool that simultaneously estimates the likelihood of having an infection due to 3GC-R, CRE, or MDRP when one or more risk factors are present among patients with infections due to Gram-negative bacteria. First, we sought to describe the risk factors for 3GC-R, CRE, and MDRP among

patients with community- and hospital-acquired Gram-negative infections using data from a large cohort of hospitalized patients across multiple facilities. Prediction models were developed from these risk factor analyses to estimate the probability of having a 3GC-R, CRE, or MDRP infection when one or more risk factors were present in a given patient. The prediction models were then used to create a user-friendly clinical instrument for use at the bedside.

In summary, our prediction tool was developed to help clinicians identify hospitalized adult patients with Gram-negative bacteria who would benefit the most from tailored empiric treatment regimens in the critical period when a Gram-negative pathogen is identified on a Gram stain or with a rapid diagnostic test and antibiotic susceptibility results are not yet available. With this information, physicians can make more informed empirical antibiotic selections, and thereby increase the likelihood of timely appropriate antibiotic therapy. Studies have shown that the critical window between infection onset and delivery of appropriate antibiotics is 48–72 h [17–19]. Our tool aids in antibiotic selection during this critical time window as Gram stain and rapid diagnostic test results become available within the first 12–24 h of culture collection.

Methods

Data source

Input data for the development of the models were from a retrospective observational study of the Premier Hospital database, which contains coding and billing information for approximately 50 million admissions from more than 500 acute-care hospitals and is the largest hospital-based database in the United States. The study was limited to the approximately 160 institutions that contributed microbiology data for the entire study period of January 1, 2011, to October 1, 2015 [20]. The database was fully de-identified and compliant with the Health Insurance Portability and Accountability Act of 1996 (HIPAA); as such, no special permission was required to review patient records and extract the data. Given the de-identified and retrospective nature of the data, as well as the observational study design, written patient consent was neither required nor sought.

Patients included in the study population were adults (≥ 18 years) who had ≥ 1 admission to a hospital with evidence of a complicated urinary tract infection (cUTI), complicated intra-abdominal infection (cIAI), hospital-acquired or ventilator-associated pneumonia (HAP/VAP), or bloodstream infection (BSI) (Additional file 2). The index culture was defined as the earliest culture drawn that produced a positive finding for any Gram-negative bacteria. In addition, patients had to have a positive culture for Gram-negative bacteria drawn from a site consistent with the infection type (Additional file 2).

Finally, patients were also required to have evidence of treatment with an intravenous antibiotic on the day of Gram-negative index culture collection or 2 days thereafter.

Infection groups were defined based on prespecified selection algorithms using primary or secondary International Classification of Diseases, Ninth Revision (ICD-9) diagnosis and procedure codes for each cohort (Additional file 2). Because codes were not mutually exclusive, an individual patient could contribute to more than one infection category during the study period.

3GC-R were defined as Enterobacteriaceae that were not susceptible to third-generation cephalosporins. CRE was defined as Enterobacteriaceae that were nonsusceptible to doripenem, meropenem, imipenem, or ertapenem. MDRP was defined as *Pseudomonas* that were not susceptible to at least three antipseudomonal agents, including penicillins, cephalosporins, monobactams, carbapenems, aminoglycosides, or fluoroquinolones. Nonsusceptibility was defined as either resistant or intermediate. Overall, susceptibility status was defined as infections that were susceptible to other antibiotics; patients who had an infection that could not be ascertained based on the susceptibility status were excluded from the study.

Potential predictors

Potential predictors in the models included infection-, patient-, and hospital-level characteristics. Additional details related to the potential predictors in the models are found in Additional file 2. Infection-level characteristics included the site of infection, type of hospital unit (intensive care unit [ICU] vs non-ICU) at time of culture draw, or hospital- or community-acquired. The infection was considered to be hospital-acquired if the patient had an index culture date ≥ 3 days after hospital admission and was considered community-acquired for patients who had an index culture date < 3 days after hospital admission.

Patient-level characteristics included age, race, sex, marital status, and a composite comorbidity index (Charlson Comorbidity Index; eg, cancer, cerebrovascular disease, chronic pulmonary disease, congestive heart failure, diabetes with or without complications, dialysis, mild liver disease, myocardial infarction, paraplegia or hemiplegia, peripheral vascular disease) [21]. Patient-level characteristics also included admission type (emergency, urgent, elective, trauma center, or other), source (transfer, clinical referral, court/law enforcement, other, or unknown), prior all-cause hospitalization during the 6-month period before the admission, infections in the 3 months prior to the admission, and prior number of antibiotics. Prior antibiotic use was defined as use of

antibiotic with activity against Gram-negative bacteria prior to index culture day in the qualified admission (Additional file 2). It was categorized as < 2 , $2-3$, and ≥ 4 , indicating cumulative number of different antibiotics a patient received before index culture date in the qualified admission.

Hospital-level characteristics included the type of facility (ie, teaching vs nonteaching), setting (ie, urban or rural), geographic area (ie, Northeast, Midwest, South, or West), geographic division (ie, New England, Middle Atlantic, East North Central, West North Central, South Atlantic, East South Central, West South Central, Mountain, or Pacific), and number of beds (ie, 0–99, 100–199, 200–299, 300–399, 400–499, and ≥ 500). Prevalence of resistant Gram-negative infections (CRE, 3GC-R, and MDRP) in the hospital facility was included and defined by the median prevalence value across all hospitals.

Statistical analyses

The first step of analysis was to summarize the study population by standard descriptive statistics (means for continuous variables, proportions for categorical variables) to compare patients with resistant and nonresistant infections. Pearson χ^2 or Fisher's exact test was performed for comparisons between two categorical variables; Student *t* or Mann-Whitney U test was used for comparisons between dichotomous and a continuous variable. Tests for trends were used to assess for the presence of a relationship between cumulative number of prior antibiotic exposures with Gram-negative activity and presence of resistant pathogen.

Because of the large number of predictors identified, least absolute shrinkage and selection operator (LASSO) logistic regression, or L1-regularized logistic regression, was used to select the set of predictors that best predicted the outcome by evaluating each coefficient simultaneously. Separate models for each resistance phenotype were conducted for hospital-acquired and community-acquired infections because they represent different patient populations with potentially different sets of risk factors for resistant infections. For the LASSO logistic regression analyses, the study sample was randomly split into the training set (70% of the study sample) and the test set (30% of the study sample). The training set was used to construct the LASSO logistic regression model. Cross-validation based on area under the curve (AUC) of receiver operating curves on training data was used to determine the best LASSO logistic regression model. Variables found to be predictive in the univariate analysis at $P < 0.20$ were included as potential predictors at model entry in each LASSO logistic regression analysis. Variables with $P < 0.1$ were retained in the final models. Model performance was evaluated using the model lift among the top 10% of scored

subjects in the test data. The model lift was defined as the probability of a positive case given a top 10% score divided by the probability of a positive case in the overall sample. A higher lift indicated a stronger association between the predicted score and the outcome. The predicted and observed likelihood of each resistant phenotype in each model was examined for any discordance.

Development of a user-friendly clinical bedside tool

Six models were developed: three for patients with hospital-acquired 3GC-R, CRE, and MDRP infections and three for community-acquired 3GC-R, CRE, and MDRP infections. A clinical prediction tool implemented using Microsoft Excel™ (Redmond, WA) was developed to provide a convenient interface to the statistical models for use at the patient bedside. Infection, hospital, and patient characteristics for an individual patient are input into the tool, which then displays the probability of a resistant infection in tabular and graphical forms. The final tool contains the hospital- and community-specific models for each category of resistant pathogens and is available from Additional file 1 together with a user guide.

Results

Study population and baseline event data

A total of 124,068 patients contributed to the dataset during the study period. Baseline event counts in the training and test datasets are shown in Additional file 2: Table S1. Numbers of patients admitted for cUTI, cIAI, HAP/VAP, and BSI were 76,367, 5649, 20,432, and 32,706, respectively (some patients contributed more than one infection type). The proportions of resistant infection in the study population overall were 12.09% for 3GC-R, 1.90% for CRE, and 3.91% for MDRP. Within each microbial category, a greater percentage of resistant infections were community-acquired relative to hospital-acquired (3GC-R, 9.27% vs of 12.09%; CRE, 1.30% vs 0.62% of 1.90% MDRP, 2.39% vs 1.59% of 3.91%).

Bivariate risk factor analysis

Detailed results of the bivariate risk factor analyses are shown in Additional file 2: Table S2. The values in the table indicate the degree of increased risk conferred by having the level of the predictor indicated for that row. For yes/no predictors, a value of “no” was taken as the reference value and indicated no additional risk. A value of 0 for a predictor indicates that the predictor was not included in that model, based on the results of the multivariate analysis. Covariates remaining in the final models were then summarized in Table 1. The number of prior antibiotics received, infection site, infection

during the previous 3 months, and hospital prevalence of the resistant pathogen were among the most important predictors across most of the six models (boldface cells in Table 1).

Predictive models for hospital-acquired gram-negative infections

Among hospital-acquired infections, the most important predictor for all three resistant phenotypes was the number of antibiotics received during the current hospital admission. Infection type was predictive of the presence of resistant infection: cUTI was not a predictor of resistance, cIAI and BSI were predictive of CRE and 3GC-R only, and HAP/VAP predicted the presence of all three resistant phenotypes. Among the infection types examined, only HAP/VAP was a predictor of MDRP. Lesser predictors of CRE and 3GC-R infection, in addition to hospital prevalence, admitting source, and hospital admission during the previous 6 months (common to both hospital- and community-acquired infection mentioned above), were patient stay in the ICU and diabetes with complications. Prior infection in the last 3 months was predictive of MDRP infection, as were chronic pulmonary disease and paraplegia/hemiplegia. Comorbidities such as cancer, cerebrovascular disease, congestive heart failure, and mild liver disease predicted the hospital acquisition of 3GC-R and to a lesser extent CRE, but not MDRP (Table 1, Additional file 2: Table S2).

Predictive models for community-acquired gram-negative infections

Predictors of all three resistant phenotypes in community-acquired infections included cUTI, infection during the 3 months before hospital admission, presence of paraplegia/hemiplegia, and diabetes without complications. For CRE, the most important predictors were prevalence, prior infection in the last 3 months, and infection site. For 3GC-R, age, prevalence, and prior infection in the last 3 months were most important. For MDRP, infection site, paraplegia/hemiplegia, prior infection in the last 3 months, and prevalence were most important (Table 1, Additional file 2: Table S2).

Evaluation of model performance

Of the six models, those for 3GC-R had the highest correct prediction rate for resistant infection for both hospital- and community-acquired infections (18.4 and 21.7%, respectively). The MDRP models had the highest AUC and the highest lift (indicating a stronger association between predicted score and outcome) for both hospital- and community-acquired infections. Because of the importance of prior antibiotic use, the hospital-acquired models had higher AUC and lift values than the corresponding community-acquired models (Table 2).

Table 1 Independent predictors of ≥ 1 type of resistant infection included in the multivariate analysis^a

Predictor	CRE		3GC-R		MDRP	
	Hospital	Community	Hospital	Community	Hospital	Community
Other	Infection site^b	✓	✓	✓	✓	✓
	ICU vs non-ICU	✓		✓		✓
	Hospital prevalence	✓	✓	✓	✓	✓
Patient	Age		✓	✓		
	Transfer	✓	✓	✓	✓	✓
	Admission in prior 6 months	✓	✓	✓	✓	✓
	Prior number of antibiotics^c	✓		✓		✓
Comorbidities ^d	Infection in prior 3 months		✓		✓	✓
	Cancer			✓		✓
	Cerebrovascular disease	✓		✓		
	Chronic pulmonary disease					✓
	Heart failure			✓		
	Diabetes with complications	✓		✓	✓	
	Diabetes without complications		✓		✓	✓
	Dialysis	✓	✓	✓	✓	✓
	Mild liver disease	✓		✓		
	Myocardial infarction	✓				✓
	Para/hemiplegia		✓		✓	✓
	Peripheral vascular disease			✓		

3GC-R third-generation cephalosporin-resistant Enterobacteriaceae, CRE carbapenem-resistant Enterobacteriaceae, ICU intensive care unit, MDRP multidrug-resistant *Pseudomonas aeruginosa*

^aBoldface cells = most important predictors across most models (predictors incurring the highest level of risk compared with the reference value).

^bComplicated urinary tract infection, complicated intra-abdominal infection, bloodstream infection, or hospital-acquired/ventilator-associated pneumonia.

^cPrior number of antibiotics was not included for community-acquired infection because of the difficulty of recovering accurate data

^dPrimary and secondary diagnoses were based on International Classification of Diseases, Ninth Revision, Clinical Modification [ICD-9-CM] codes

Creation of a user-friendly Excel-based clinical bedside tool

To enable application of the six predictive multivariate logistic regression models to real-world clinical practice, we developed a user-friendly interface that estimates the risk of resistant infection. This tool is intended for use by clinicians at the patient bedside. Because it is not intended to be prescriptive, the tool simply estimates the risk of resistance without making

specific recommendations regarding the best treatment. These recommendations will help the clinician select the most appropriate (empiric) antibiotic treatment for an individual patient according to the full context of patient characteristics and circumstances.

Consisting of four worksheets, the full tool and a detailed user guide are available at Additional file 1. User-modifiable drop-down lists are provided for entry of the model inputs of infection, hospital, and patient

Table 2 Evaluation of model predictive performance

Performance Metric ^a	CRE		3GC-R		MDRP	
	Hospital	Community	Hospital	Community	Hospital	Community
AUC for training data	0.90	0.79	0.87	0.71	0.94	0.84
AUC for test data	0.92	0.79	0.87	0.70	0.94	0.83
Correct prediction among top 10% scored subjects in test data, n/N (%)	145/3496 (4.15)	165/3607 (4.57)	627/3411 (18.38)	786/3624 (21.69)	405/3586 (11.29)	388/3628 (10.69)
Lift of top 10% scored subjects in test data ^b	7.8	3.8	6.4	2.6	7.9	5.0

3GC-R third-generation cephalosporin-resistant Enterobacteriaceae, AUC area under the receiver operating characteristic curve, CRE carbapenem-resistant Enterobacteriaceae, LASSO least absolute shrinkage and selection operator, MDRP multidrug-resistant *Pseudomonas aeruginosa*

^aThe method of predictor selection was LASSO logistic regression that minimized the cross-validation misclassification error; LASSO C value was 0.1

^bLift defined as probability of a positive case given a top 10% score divided by the probability of a positive case in overall sample. This ratio evaluates how much a top score enriches for selecting positive cases compared with random sampling in the absence of a model. A higher lift indicates a stronger association between the predicted score and the outcome

characteristics. A summary of the results is then provided, consisting of the risk factors and the likelihood of the presence of each type of resistant infection in the model, including the number of patients that need to be observed to detect one case of resistance (1/probability), which is easier to interpret in cases of small probabilities. A summary of the results for each run of the tool can be printed as a portable document format (PDF) report that includes the tool overview, model inputs, and results. In addition, a comprehensive calculation sheet can be accessed if required for more detailed data analysis (Fig. 1).

Discussion

One of the fundamental pillars of antimicrobial stewardship is ensuring that patients with life-threatening infections receive early, appropriate antimicrobial therapy. Despite the longstanding recognition of the positive benefits of “getting it right the first time,” delayed appropriate therapy rates for patients with serious Gram-negative infections are still reported to be > 30% in several publications [3, 7]. To facilitate the administration of early appropriate therapy, it is important that patients at risk of being infected with resistant pathogens be identified

promptly, especially as definitive culture results are typically not available within the first 24 to 72 h of onset of infection [15, 16].

Advances in rapid diagnostics have shortened the lag time between infection onset to pathogen identification from days to hours and have had positive effects on clinical outcomes when paired with robust antimicrobial stewardship programs [22]. Although rapid diagnostics represent a significant advance from traditional culture methods, current technologies are only able to identify a limited number of antibiotic-resistant Gram-negative pathogens. Therefore, clinical prediction tools, ideally in tandem with rapid diagnostic tests and Gram stain results, are needed to inform empiric antibiotic selection in the critical period when a Gram-negative pathogen is identified on a Gram stain or with a rapid diagnostic test and antibiotic susceptibility results are not yet available. To date, most published clinical prediction tools have focused on one pathogen or antibiotic-resistant phenotype [23, 24]. Although helpful in the antibiotic selection process, the risk factors for infections due to the various antibiotic-resistant Gram-negative pathogens are largely overlapping, and patients are at risk of several resistant Gram-negative pathogens simultaneously when a

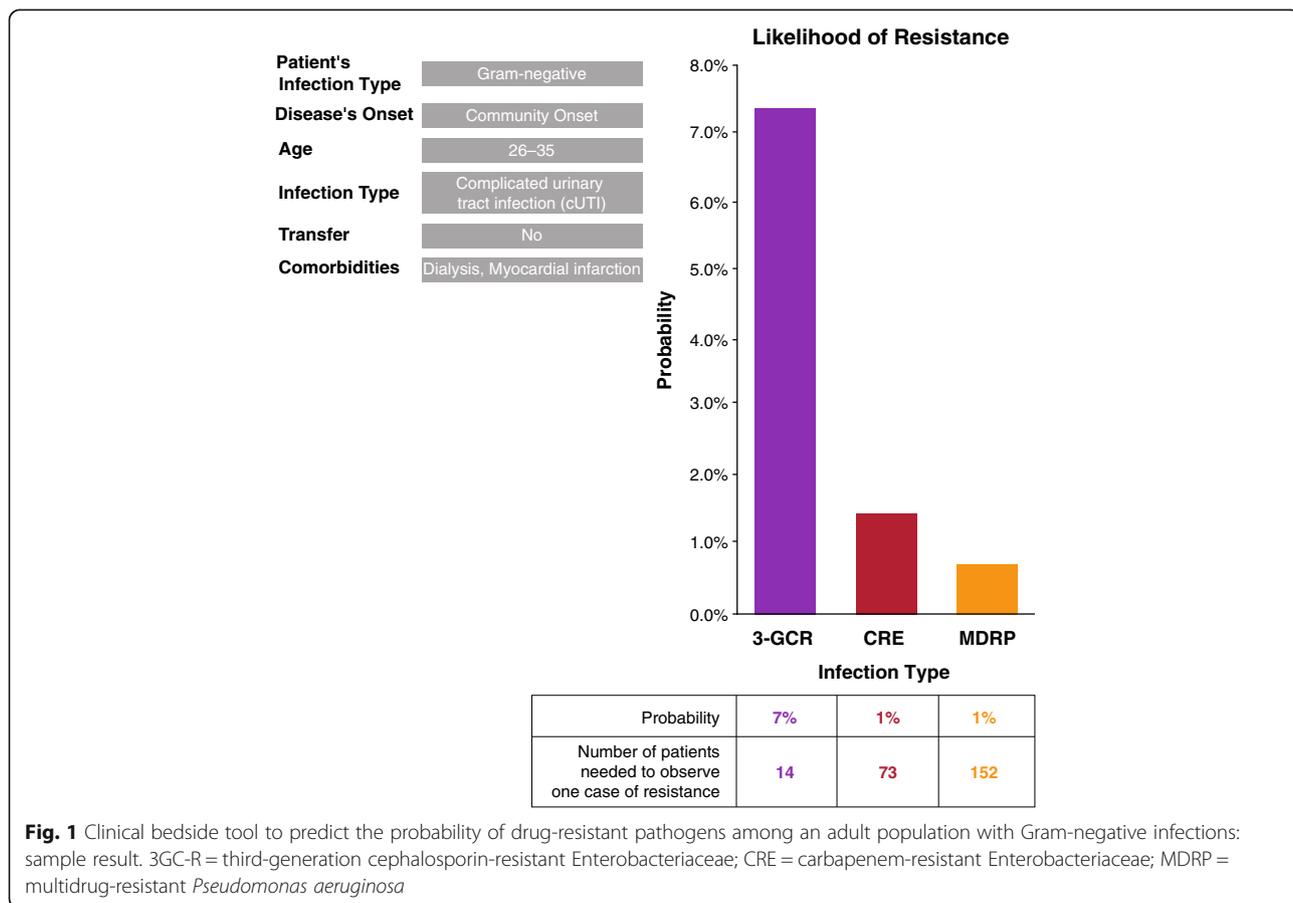


Fig. 1 Clinical bedside tool to predict the probability of drug-resistant pathogens among an adult population with Gram-negative infections: sample result. 3GC-R = third-generation cephalosporin-resistant Enterobacteriaceae; CRE = carbapenem-resistant Enterobacteriaceae; MDRP = multidrug-resistant *Pseudomonas aeruginosa*

common risk factor, or combination of risk factors, are present. For example, prior receipt of carbapenems has been found to increase the risk of having an infection due to several highly resistant Gram-negative bacteria. Cognizant of this, we developed a clinical prediction tool that estimates the probabilities of having a Gram-negative infection due to 3GC-R, CRE, or MDRP. We selected these antibiotic-resistant Gram-negative pathogens because they are becoming increasingly common across most US healthcare institutions and are considered major threats to public health by the US Centers for Disease Control and Prevention [1].

When developing the clinical tool, we also believed it was important to develop two different models to differentiate the two distinct hospitalized patient populations with Gram-negative infections because risk factors or strength of their association can vary by population. The first focused on patients who presented to the hospital (ie, community-acquired); the second centered on patients who developed their infection during their hospitalization (ie, hospital-acquired). Interestingly, patients with community-acquired Gram-negative infections had a higher baseline risk of having resistant Gram-negative infections relative to those with hospital-acquired infections in this study, highlighting the need for the development of two separate clinical prediction models. Another distinguishing feature of this clinical prediction model was the inclusion of the prevalence of each resistant phenotype of interest in the hospital where the patient developed the infection. It is well established that a patient's risk of having an antibiotic-resistant infection is driven in part by the bacteria present in the healthcare institution. Therefore, we believe it was important to consider it as a covariate in the clinical model-building phase. Finally, we believe that our model also rectifies issues related to autocorrelation, in which a patient could theoretically be counted twice if exposed to more than one type of infection. This exposure could potentially occur on different sites or on different days, leading to potentially different exposures. Because we considered the site of infection in model development, the risk of autocorrelation was minimized, confirming the quality of our model.

Although there were some distinguishing aspects to our clinical prediction tool, our model development approach was similar to previous studies [25–27]. Using the Premier Hospital database, which included 124,068 hospital admissions from approximately 160 institutions that contributed microbiology data during the period from January 1, 2011, to October 1, 2015, we first identified the infection-, patient-, and hospital-level risk factors that increase the probability of having an infection due to 3GC-R, CRE, or MDRP. Not surprisingly, risk factors and exposures associated with antimicrobial

resistance in this study were largely consistent with previous reports [28–35]. The most important independent risk factors for both hospital and community acquisition of all three resistant bacteria were hospital prevalence of the resistant pathogen, admission source, and previous hospital admission within the prior 6 months [28–30, 32–35]. All three of these factors capture time at risk in healthcare facilities among patient populations predisposed to infection by antibiotic-resistant pathogens [28–35] and highlight the importance of understanding the prevalence of a given antibiotic-resistant phenotype in an institution. Another common risk factor for 3GC-R, CRE, and MDRP infection among individuals with hospital-acquired Gram-negative infections was the number of antibiotics received during the current admission [28–35]. Constant and cumulative exposure to antibiotics disturbs the natural bacterial flora, in particular in the gastrointestinal tract, and predisposes patients to colonization by resistant phenotypes. For all three pathogens, the presence of a resistant phenotype was predicted more strongly by previous use of four or more antibiotics than by previous use of two to three antibiotics in the current hospital admission. Our data suggest that a patient's cumulative antibiotic exposure history is likely to be more important than any one specific antibiotic exposure when determining a patient's likelihood of harboring a resistant pathogen.

The information from the multivariate logistic regression analyses was then used to develop models to estimate the likelihood of having these infections when one or more risk factors were present in hospitalized adult patients with Gram-negative infections. The major advantage of using logistic regression to develop clinical prediction rules is the functionality of the final models. In addition to identifying variables that are independently associated with stronger odds of having the outcome of interest, the final logistic regression models are mathematic equations that can be used to predict the probability of antimicrobial resistance based on the combination of significant risk factors present in a given individual with an infection [10, 36]. Adaptation of the models to provide a clinical tool was relatively straightforward because of their simplicity. With the information generated from this clinical prediction tool, we anticipate that clinicians will be able to make more informed empiric antibiotic selection decisions and thereby increase the likelihood of appropriate empiric antibiotic therapy. Although no specific recommendations are made regarding treatment options, the tool is designed as a simple interface to estimate the risk of resistance, which can be used by the clinician to determine the best course of treatment at bedside.

Several things should be considered when interpreting these findings. As the data used for the development of

our models was from a database, our study is subject to the limitations associated with retrospective observations studies, and the ICD-9 codes may not be 100% accurate. As with all electronic health databases, there may be errors of omission and/or commission in coding. Because our operational definitions were based on information within the database, study measures may be less accurate than those based on medical record review or data gathered prospectively. Because the Premier Hospital database lacks information on healthcare utilization outside of Premier facilities, we did not include prior receipt of antibiotics in the community-acquired model. We did not consider prior colonization with a resistant pathogen as part of these analyses as only clinical culture data were available in the database. We also cannot exclude the possibility of patient-to-patient transmission of the resistant strains, which may weaken the association between acquisition of resistant pathogens and the identified risk factors. Another limitation was that the tool was not validated using an external dataset; a validation study should be performed. Finally, additional prediction methods such as neural networks, random forest, and SuperLearner, which allow for the incorporation of several algorithms simultaneously to deliver the strongest prediction model, may improve the prediction modeling observed in this study [37–39]. Despite these limitations, we think the model fit statistics demonstrate that we employed robust methodologies to derive the clinical prediction tool and adequately captured comorbid conditions, key baseline characteristics, and clinical covariates when deriving clinical prediction tools. More importantly, we believe our clinical prediction tool has merit, as it relied on the data elements that are typically available to the clinician at the time of empiric antibiotic selection among patients presenting with Gram-negative infections.

Conclusions

Based on a large retrospective observational study, we developed six separate models for the prediction of hospital- and community-acquired infections due to 3GC-R, CRE, and MDRP among hospitalized adult patients with Gram-negative infections. The performance of our models is superior to or comparable with the performance of similar published models because of (1) the large number of patients and institutions contributing data, (2) the number and diversity of potential predictors considered, and (3) the inclusion of antibiotic resistance rates in the included hospitals. Our predictive model was implemented as a user-friendly bedside tool for use by physicians or healthcare professionals to predict the probability of resistant infection in an individual patient to expedite and direct initial antibiotic therapy and improve outcomes among hospitalized adult patients with

Gram-negative infections in the critical period when a Gram-negative pathogen is identified on a Gram-stain or with a rapid diagnostic test and antibiotic susceptibility results are not yet available.

Additional files

Additional file 1: contains the Clinical Bedside Tool. (XLSM 162 kb)

Additional file 2: contains Appendices A, B, and C, Tables S1, S2, and S3. (DOCX 51 kb)

Abbreviations

3GC-R: third-generation cephalosporin-resistant Enterobacteriaceae; AUC: area under the curve; BSI: bloodstream infection; cIAI: complicated intra-abdominal infection; CRE: carbapenem-resistant Enterobacteriaceae; cUTI: complicated urinary tract infection; HAP: hospital-acquired pneumonia; HIPAA: Health Insurance Portability and Accountability Act of 1996; ICD-9: International Classification of Diseases, Ninth Revision; ICU: intensive care unit; LASSO: least absolute shrinkage and selection operator; MDRP: multidrug-resistant *Pseudomonas aeruginosa*; ROC: receiver operating curves; VAP: ventilator-associated pneumonia

Acknowledgments

Editorial support for development of this manuscript was provided by Moira A. Hudson, PhD, CMPP, Jennifer L. Venzie, PhD, Todd J. Waldron, PhD, and John E. Fincke, PhD, at Complete Healthcare Communications, LLC (North Wales, PA), a CHC Group company. Portions of this manuscript were previously published as a poster at IDWeek 2018, October 3–7, 2018, San Francisco, CA, USA.

Author contributions

TL, NGB, JMY, HJF, and PG made substantial contributions to the study conception and design. TL, NGB, JMY, HJF, and PG collected, analyzed, and interpreted the data. TL, NGB, JMY, HJF, and PG were involved in the drafting of the manuscript and approved the submission of the manuscript for publication. TL, NGB, JMY, HJF, and PG are accountable for all aspects of the work, including the accuracy and integrity of the manuscript. All authors have read and approved the manuscript.

Authors' information

Not applicable

Funding

This work was supported by Allergan plc. Neither honoraria nor payments were made for authorship. Allergan sponsored the study and funded medical writing support. Because three authors are Allergan employees, Allergan played a role in the design of the study; the collection, analysis, and interpretation of data; and in writing the manuscript.

Availability of data and materials

The data that support the findings of this study are available from Premier Hospital, but restrictions apply to the availability of these data, which were used under license for the current study and so are not publicly available. However, data are available from the authors upon reasonable request and with permission of Premier Hospital.

Ethics approval and consent to participate

Not applicable; the database is fully de-identified and compliant with the Health Insurance Portability and Accountability Act of 1996 (HIPAA). Given the de-identified and retrospective nature of the data and the noninterventional study design, written patient consent was neither required nor sought. Administrative permissions were not required to access the raw data.

Consent for publication

Not applicable; individual patient data are not presented.

Competing interests

All authors met the International Committee of Medical Journal Editors (ICMJE) authorship criteria. Neither honoraria nor payments were made for authorship. TL has received consulting fees or honoraria from Allergan plc. He has also been a consultant for Merck and The Medicines Company and has received payment for lectures, including service on speaker bureaus for Allergan for work not associated with the current study. NGB, JMY, and PG are employees of Allergan. HJF is an employee of Evidera, a healthcare consulting and contract research firm. In his salaried positions, he is precluded from receiving payment or honoraria directly from organizations for services rendered. Evidera has received funding for the study from Allergan and has received funding from many other biomedical companies in support of various research studies and projects.

Author details

¹Albany College of Pharmacy and Health Sciences, Albany, NY 12208-3492, USA. ²Allergan plc, Irvine, CA, USA. ³Allergan plc, Madison, NJ, USA. ⁴Evidera, San Francisco, CA, USA.

Received: 19 October 2018 Accepted: 6 August 2019

Published online: 14 August 2019

References

- Centers for Disease Control and Prevention. Antibiotic resistance threats in the United States, 2013. Available at: <http://www.cdc.gov/drugresistance/threat-report-2013/pdf/ar-threats-2013-508.pdf>. Accessed 8 Aug 2019.
- Taccconelli E, Carrara E, Savoldi A, Harbarth S, Mendelson M, Monnet DL, Pulcini C, Kahlmeter G, Kluytmans J, Carmeli Y, et al. Discovery, research, and development of new antibiotics: the WHO priority list of antibiotic-resistant bacteria and tuberculosis. *Lancet Infect Dis*. 2018;18:318–27.
- Lodise TP Jr, Patel N, Kwa A, Graves J, Furuno JP, Graffunder E, Lomaestro B, McGregor JC. Predictors of 30-day mortality among patients with *Pseudomonas aeruginosa* bloodstream infections: impact of delayed appropriate antibiotic selection. *Antimicrob Agents Chemother*. 2007;51:3510–5.
- Sturkenboom MC, Goettsch WG, Picelli G, in't veld B, Yin DD, de Jong RB, Go PM, Herings RM. Inappropriate initial treatment of secondary intra-abdominal infections leads to increased risk of clinical failure and costs. *Br J Clin Pharmacol*. 2005;60:438–43.
- Tellado JM, Sen SS, Caloto MT, Kumar RN, Nocea G. Consequences of inappropriate initial empiric parenteral antibiotic therapy among patients with community-acquired intra-abdominal infections in Spain. *Scand J Infect Dis*. 2007;39:947–55.
- Raman G, Avendano E, Berger S, Menon V. Appropriate initial antibiotic therapy in hospitalized patients with gram-negative infections: systematic review and meta-analysis. *BMC Infect Dis*. 2015;15:395.
- Iregui M, Ward S, Sherman G, Fraser VJ, Kollef MH. Clinical importance of delays in the initiation of appropriate antibiotic treatment for ventilator-associated pneumonia. *Chest*. 2002;122:262–8.
- Bonine NG, Berger A, Altincatal A, Wang R, Bhagnani T, Gillard P, Lodise T. Associations between timeliness of therapy and clinical and economic outcomes among patients with serious infections due to gram-negative bacteria (GNB): how much does delayed appropriate therapy (DAT) matter? *Open Forum Infect Dis*. 2017;4:5283–4.
- Zilberberg MD, Nathanson BH, Sulham K, Fan W, Shorr AF. Carbapenem resistance, inappropriate empiric treatment and outcomes among patients hospitalized with Enterobacteriaceae urinary tract infection, pneumonia and sepsis. *BMC Infect Dis*. 2017;17:279.
- Lodise TP, McKinnon PS, Swiderski L, Rybak MJ. Outcomes analysis of delayed antibiotic treatment for hospital-acquired *Staphylococcus aureus* bacteremia. *Clin Infect Dis*. 2003;36:1418–23.
- Friedman ND, Temkin E, Carmeli Y. The negative impact of antibiotic resistance. *Clin Microbiol Infect*. 2016;22:416–22.
- Infectious Diseases Society of America, Spellberg B, Blaser M, Guidos RJ, Boucher HW, Bradley JS, Eisenstein BI, Gerding D, Lynfield R, Reller LB, et al. Combating antimicrobial resistance: policy recommendations to save lives. *Clin Infect Dis*. 2011;52:S397–428.
- Dellit TH, Owens RC, McGowan JE, Gerding DN, Weinstein RA, Burke JP, Huskins WC, Paterson DL, Fishman NO, Carpenter CF, et al. Infectious Diseases Society of America and the Society for Healthcare Epidemiology of America guidelines for developing an institutional program to enhance antimicrobial stewardship. *Clin Infect Dis*. 2007;44:159–77.
- World Health Organization. Global Action Plan on Antimicrobial Resistance. Available at: <http://www.who.int/antimicrobial-resistance/global-action-plan/en/>. Accessed 8 Aug 2019.
- Kollef MH, Sherman G, Ward S, Fraser VJ. Inadequate antimicrobial treatment of infections: a risk factor for hospital mortality among critically ill patients. *Chest*. 1999;115:462–74.
- Leibovici L, Shraga I, Drucker M, Konigsberger H, Samra Z, Pitlik SD. The benefit of appropriate empirical antibiotic treatment in patients with bloodstream infection. *J Intern Med*. 1998;244:379–86.
- Lodise TPJ, Patel N, Kwa A, Graves J, Furuno JP, Graffunder E, Lomaestro B, McGregor JC. Predictors of 30-day mortality among patients with *Pseudomonas aeruginosa* bloodstream infections: impact of delayed appropriate antibiotic selection. *Antimicrob Agents Chemother*. 2007;51:3510–5.
- Lodise TP, Zhao Q, Fahrback K, Gillard PJ, Martin A. A systematic review of the association between delayed appropriate therapy and mortality among patients hospitalized with infections due to *Klebsiella pneumoniae* or *Escherichia coli*: how long is too long? *BMC Infect Dis*. 2018;18:625.
- Bonine NG, Berger A, Altincatal A, Wang R, Bhagnani T, Gillard P, Lodise T. Impact of delayed appropriate antibiotic therapy on patient outcomes by antibiotic resistance status from serious gram-negative bacterial infections. *Am J Med Sci*. 2019;357:103–10.
- Premier Life Sciences. Premier Healthcare Database: Data that Informs and Performs. Premier Inc. Available at: <https://www.premierinc.com/downloads/PremierHealthcareDatabaseWhitepaper.pdf>. Accessed 8 Aug 2019.
- Charlson ME, Pompei P, Ales KL, MacKenzie CR. A new method of classifying prognostic comorbidity in longitudinal studies: development and validation. *J Chronic Dis*. 1987;40:373–83.
- Edmiston CE, Garcia R, Barnden M, DeBaun B, Johnson HB. Rapid diagnostics for bloodstream infections: a primer for infection preventionists. *Am J Infect Control*. 2018;46:1060–8.
- Miller BM, Johnson SW. Demographic and infection characteristics of patients with carbapenem-resistant Enterobacteriaceae in a community hospital: development of a bedside clinical score for risk assessment. *Am J Infect Control*. 2016;44:134–7.
- Jaimes F, Arango C, Ruiz G, Cuervo J, Botero J, Velez G, Upegui N, Machado F. Predicting bacteremia at the bedside. *Clin Infect Dis*. 2004;38:357–62.
- Mariscalco G, Biancarfi F, Zanobini M, Cottini M, Piffaretti G, Saccocci M, Banach M, Beghi C, Angelini GD. Bedside tool for predicting the risk of postoperative atrial fibrillation after cardiac surgery: the POAF score. *J Am Heart Assoc*. 2014;3:e000752.
- Mehta RH, Grab JD, O'Brien SM, Bridges CR, Gammie JS, Haan CK, Ferguson TB, Peterson ED. Bedside tool for predicting the risk of postoperative dialysis in patients undergoing cardiac surgery. *Circulation*. 2006;114:2208–16 quiz.
- Vasudevan A, Mukhopadhyay A, Li J, Yuen EG, Tambyah PA. A prediction tool for nosocomial multi-drug resistant gram-negative bacilli infections in critically ill patients - prospective observational study. *BMC Infect Dis*. 2014;14:615.
- Bilavsky E, Temkin E, Lerman Y, Rabinovich A, Salomon J, Lawrence C, Rossini A, Salvia A, Samsó JV, Fierro J, et al. Risk factors for colonization with extended-spectrum beta-lactamase-producing Enterobacteriaceae on admission to rehabilitation centres. *Clin Microbiol Infect*. 2014;20:0804–10.
- Denis B, Lafaurie M, Donay JL, Fontaine JP, Oksenhendler E, Raffoux E, Hennequin C, Allez M, Socie G, Maziers N, et al. Prevalence, risk factors, and impact on clinical outcome of extended-spectrum beta-lactamase-producing *Escherichia coli* bacteraemia: a five-year study. *Int J Infect Dis*. 2015;39:1–6.
- Gasink LB, Edelstein PH, Lautenbach E, Synnestvedt M, Fishman NO. Risk factors and clinical impact of *Klebsiella pneumoniae* carbapenemase-producing *K. pneumoniae*. *Infect Control Hosp Epidemiol*. 2009;30:1180–5.
- Harris AD, McGregor JC, Johnson JA, Strauss SM, Moore AC, Standiford HC, Hebden JN, Morris JG Jr. Risk factors for colonization with extended-spectrum beta-lactamase-producing bacteria and intensive care unit admission. *Emerg Infect Dis*. 2007;13:1144–9.
- Zarkotou O, Pournaras S, Tselioti P, Dragoumanos V, Pitiiriga V, Ranellou K, Prekates A, Themeli-Digalaki K, Tsakris A. Predictors of mortality in patients with bloodstream infections caused by KPC-producing *Klebsiella pneumoniae* and impact of appropriate antimicrobial treatment. *Clin Microbiol Infect*. 2011;17:1798–803.

33. Sullivan T, Ichikawa O, Dudley J, Li L, Aberg J. The rapid prediction of carbapenem resistance in patients with *Klebsiella pneumoniae* bacteremia using electronic medical record data. *Open Forum Infect Dis*. 2018;5:ofy091.
34. Goodman KE, Lessler J, Cosgrove SE, Harris AD, Lautenbach E, Han JH, Milstone AM, Massey CJ, Tamma PD. A clinical decision tree to predict whether a bacteremic patient is infected with an extended-spectrum beta-lactamase-producing organism. *Clin Infect Dis*. 2016;63:896–903.
35. Yu Y, Shen H, Zhu C, Guo R, Gao Y, Lu L. Infections caused by extended-spectrum beta-lactamase producing *Escherichia coli* in systemic lupus erythematosus patients: prevalence, risk factors, and predictive model. *Biomed Res Int*. 2018;2018:8296720.
36. Lodise TP, Miller CD, Graves J, Furuno JP, McGregor JC, Lomaestro B, Graffunder E, McNutt LA. Clinical prediction tool to identify patients with *Pseudomonas aeruginosa* respiratory tract infections at greatest risk for multidrug resistance. *Antimicrob Agents Chemother*. 2007;51:417–22.
37. Rose S. Mortality risk score prediction in an elderly population using machine learning. *Am J Epidemiol*. 2013;177:443–52.
38. Li Y, Wu FX, Ngom A. A review on machine learning principles for multi-view biological data integration. *Brief Bioinform*. 2018;19:325–40.
39. Degenhardt F, Seifert S, Szymczak S. Evaluation of variable selection methods for random forests and omics data sets. *Brief Bioinform*. 2019;20:492–503.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Ready to submit your research? Choose BMC and benefit from:

- fast, convenient online submission
- thorough peer review by experienced researchers in your field
- rapid publication on acceptance
- support for research data, including large and complex data types
- gold Open Access which fosters wider collaboration and increased citations
- maximum visibility for your research: over 100M website views per year

At BMC, research is always in progress.

Learn more biomedcentral.com/submissions

